

## DETERMINATION OF THE RATE CONSTANT OF THE SOLVENT CAGE REORIENTATION AROUND A PHOTOEXCITED MOLECULE USING PHASE MODULATION FLUOROMETRY

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### Summary

The fluorescence lifetime and quantum yield of ethylamino-9-methoxy-2-chloro-6-acridine in glycerol solution were measured at three temperatures using the phase modulation technique. The results were interpreted in terms of a relaxation process in the excited state that involved two emitting species: one (designated F) directly excited and one (designated R) resulting from the reorientation of the solvent cage around the F state. The rate constant of this process was evaluated; it was greatly increased on raising the temperature, which also enhanced the radiationless deactivation rate constants of both emitting species, although to a minor extent. At 60 °C, this relaxation process is so fast that the whole fluorescence is emitted from R species.

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### 1. Introduction

9-aminoacridinic compounds (9AA) are widely used as fluorescence probes in biological fields. Our aim was to study the change of their fluorescence properties in solution under various experimental conditions (viscosity, temperature, pH, solvent used etc.). A previously published study showed that a solvent relaxation process occurs after the photoexcitation of molecules of 9AA [1]. This process can be investigated by performing phase modulation fluorescence measurements if it occurs at a rate that is of the same order of magnitude as the rates of other deactivation mechanisms from the excited state [2, 3]. The results may then be interpreted by using the two excited state (F and R) model of Lakowicz and Balter [4]. In our case, the directly photoexcited F state would be solvated as in the ground state while the excited state R would result from a different orientation of solvent molecules around the excited state F (Fig. 1).

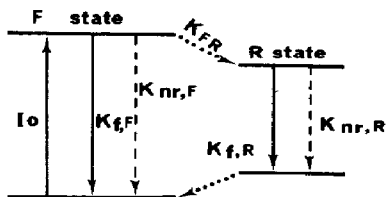


Fig. 1. Energy diagram of F and R states:  $I_0$  is the number of photons absorbed per second;  $k_{f,F}$  and  $k_{f,R}$  are the rate constants of radiative transitions from the F and R states;  $k_{FR}$  is the rate constant of the F  $\rightarrow$  R transition;  $k_{nr,F}$  and  $k_{nr,R}$  are the rate constants of other radiationless transitions from F and R states.

In this work we have studied 9-ethylamino-2-methoxy-6-chloroacridine (ECMA) in glycerol solutions at various temperatures. Our purpose was to look for any influence of the viscosity of the medium and the temperature on the solvent relaxation rate constant  $k_{FR}$ . We determined  $k_{FR}$  from phase modulation and quantum yield measurements.

## 2. Experimental details

ECMA was synthesized at the laboratory of Dr. P. Jacquignon (Institut des Substances Naturelles, Gif sur Yvette, France). Fluorescence spectra were recorded using a Jobin-Yvon 3 D apparatus. Absorbance measurements were performed using a Uvikon 820 spectrophotometer. Fluorescence quantum yields were determined by comparison with a standard of known fluorescence quantum yield [5]. The standard was a solution of atebtrin in methanol with a quantum yield of 0.111 [6].

Phase shift and demodulation factor measurements were performed using an SLM 4800 spectrofluorometer. The apparent phase fluorescence lifetime  $\tau_\phi$  was determined from the phase difference  $\phi$  between the fluorescence and the exciting light modulated at the angular frequency  $\omega = 2\pi\nu$  according to

$$\tau_\phi = \frac{1}{\omega} \tan \phi \quad (1)$$

The apparent modulation fluorescence lifetimes  $\tau_m$  were determined from the demodulation factor  $m$  according to the relation

$$\tau_m = \frac{1}{\omega} \left( \frac{1}{m^2} - 1 \right)^{1/2} \quad (2)$$

Each  $\tau_\phi$  and  $\tau_m$  represent the average of a set of ten measurements, each of which, in turn, was the average of a set of 100 separate values obtained at a sampling rate of approximately  $5 \text{ s}^{-1}$ . The maximum coefficient of variation in an independent measurement of either  $\tau_\phi$  or  $\tau_m$  was 5%. Data were corrected for the wavelength-time response of the photomultiplier tube accord-

ing to the method of Jameson and Weber [7]. Consequently, an error treatment was rather difficult to perform.

When the coefficient of variation between the  $\tau_\phi$  and the  $\tau_m$  reached 20%, we considered that these parameters had different values which were dependent on the experimental conditions. However, when this coefficient was only 6%,  $\tau_\phi$  and  $\tau_m$  were considered to be equal whatever the experimental conditions.

Thus, our interpretation relied not only on the eventual difference between  $\tau_\phi$  and  $\tau_m$  measured under the same experimental conditions but also on the change in both these parameters with angular frequency or observation wavelength. Control measurements were performed according to the technique of Spencer and Weber [8] to confirm that the rotational Brownian motion of the molecules had no influence on the experimental data.

### 3. Results

#### 3.1. Phase modulation experiment

The data from phase modulation measurements of a  $2.5 \times 10^{-5}$  M ECMA-glycerol solution at 5 °C are listed in Table 1. It can be seen that for short wavelengths  $\tau_\phi < \tau_m$ ,  $m/\cos \phi < 1$  and  $\tau_\phi$ ,  $\tau_m$  and  $m/\cos \phi$  were decreasing functions of  $\omega$ ; for long wavelengths  $\tau_\phi > \tau_m$ ,  $m/\cos \phi > 1$  and  $\tau_\phi$ ,  $\tau_m$  and  $m/\cos \phi$  were increasing functions of  $\omega$ . At a wavelength close to 530 nm,  $\tau_\phi$  and  $\tau_m$  were equal whatever the modulation frequency. In addition,  $\tau_\phi$  and  $\tau_m$  increased with  $\lambda$  over the whole wavelength range studied. All these results are typical of an excited state process occurring with a rate constant of the same order of magnitude as those of other deactivation processes from this excited state [2 - 4]. This excited state process may be due to the reorientation of solvent molecules around photo-excited ECMA molecules, as is the case for other similar compounds [3].

TABLE 1

Phase modulation fluorescence measurements on a  $2.5 \times 10^{-5}$  M ECMA-glycerol solution at 5 °C

$\lambda_{\text{obs}}$ (nm)	30 MHz			18 MHz			6 MHz		
	$\tau_\phi$	$\tau_m$	$m/\cos \phi$	$\tau_\phi$	$\tau_m$	$m/\cos \phi$	$\tau_\phi$	$\tau_m$	$m/\cos \phi$
460	5.6	8.3	0.78	7.2	10.1	0.85	9.2	12.7	0.95
480	9.5	12.6	0.80	10.8	13.2	0.88	13.8	13.9	1.00
500	10.9	14.0	0.81	13.6	14.6	0.95	14.0	14.4	0.99
520	13.2	14.6	0.92	14.6	15.2	0.97	14.8	15.0	1.00
540	16.7	15.3	1.09	16.7	15.5	1.04	15.6	15.5	1.00
560	18.1	16.1	1.08	17.9	15.7	1.04	15.7	16.2	0.99
575	21.6	16.3	1.30	19.5	16.0	1.12	16.3	16.0	1.01

The fluorescence was then emitted from both F and R states (Fig. 1) and  $\tau_\phi$  and  $\tau_m$  may be expressed as [9]

$$\tau_\phi = \frac{\alpha\tau_F(1 + \omega^2\tau_R^2) + (1 - \alpha)(\tau_F + \tau_R)}{\alpha(1 + \omega^2\tau_R^2) + (1 - \alpha)(1 - \omega^2\tau_F\tau_R)} \quad (3)$$

$$\tau_m = \left( \frac{\tau_F^2\tau_R^2\omega^2 + \tau_F^2 + \tau_R^2 - \alpha^2\tau_R^2}{1 + \alpha^2\tau_R^2\omega^2} \right)^{1/2} \quad (4)$$

In eqns. (3) and (4)  $\tau_F$  and  $\tau_R$  are the lifetimes of the excited states F and R and  $\alpha$  is the contribution of the F state to the overall fluorescence.

$\tau_R$  was determined as  $15.0 \pm 0.5$  ns, *i.e.* the mean value of  $\tau_\phi$  and  $\tau_m$  obtained by interpolation at  $\lambda = 530$  nm where  $\tau_\phi$  is close to  $\tau_m$  (and  $m/\cos \phi$  is close to unity) and  $\tau_\phi$  and  $\tau_m$  are independent of modulation frequency. A theoretical study of the variations in  $\tau_\phi$  and  $\tau_m$  with  $\alpha$  showed that in this case  $\alpha = \tau_F/\tau_R$  and eqns. (3) and (4) reduce to  $\tau_\phi = \tau_m = \tau_R$  [3, 9]. Using  $\tau_R = 15.0$  ns,  $\tau_F$  and  $\alpha$  can be calculated from eqns. (3) and (4) for each observation wavelength and modulation frequency. The results are listed in Table 2.

TABLE 2

$\tau_F$  and  $\alpha$  determined at different observation wavelengths for an ECMA-glycerol solution at  $\theta = 5^\circ\text{C}$  and  $\theta = 21^\circ\text{C}$

		$\lambda$ (nm)						
		460	480	500	520	540	560	575
$\theta = 5^\circ\text{C}$								
30 MHz	$\tau_F$ (ns)	4.0	4.0	3.0	3.0	4.0	4.5	4.5
	$\alpha$	0.66	0.38	0.34	0.30	0.26	0.23	0.22
18 MHz	$\tau_F$ (ns)	4.0	4.5	4.0	—	5.0	4.5	4.5
	$\alpha$	0.59	0.39	0.31	—	0.24	0.22	0.20
$\theta = 21^\circ\text{C}$								
30 MHz	$\tau_F$ (ns)	1.5	1.5	0.8	—	1.5	1.2	1.3
	$\alpha$	0.33	0.16	0.14	0.10	0.09	0.08	0.07
18 MHz	$\tau_F$ (ns)	1.7	2.5	—	2.0	1.5	—	1.0
	$\alpha$	0.32	0.16	—	0.09	0.08	—	0.06

Experiments were performed at  $21^\circ\text{C}$  and  $60^\circ\text{C}$  to determine the influence of temperature and viscosity on the rate constant  $k_{FR}$ .

At  $21^\circ\text{C}$ , the relaxation process in the excited state was still apparent, as shown by the changes in  $\tau_\phi$  and  $\tau_m$  with  $\omega$  and  $\lambda$ , and their relative values (Table 3). Nevertheless,  $\tau_F$  was shorter:  $\tau_F = 1.5 \pm 0.5$  ns at  $21^\circ\text{C}$  compared with  $\tau_F = 4.1 \pm 0.6$  ns at  $5^\circ\text{C}$ ; in addition the  $\alpha$  were also smaller (Table 2).

At  $60^\circ\text{C}$ , the excited state process was no longer apparent.  $\tau_\phi$  and  $\tau_m$  could be considered equal within experimental error and were independent of observation wavelength and modulation frequency used (Table 4). Thus,

TABLE 3

Phase modulation fluorescence measurements on a  $2.5 \times 10^{-5}$  M ECMA-glycerol solution at 21 °C

$\lambda_{\text{obs}}$ (nm)	30 MHz			18 MHz			6 MHz		
	$\tau_{\phi}$	$\tau_m$	$m/\cos \phi$	$\tau_{\phi}$	$\tau_m$	$m/\cos \phi$	$\tau_{\phi}$	$\tau_m$	$m/\cos \phi$
460	5.1	10.7	0.62	7.6	12.2	0.77	9.9	13.5	0.95
480	9.9	13.5	0.77	12.1	13.7	0.92	12.5	13.6	0.98
500	11.2	14.2	0.82	13.4	14.4	0.95	12.9	11.9	1.01
520	14.7	14.4	1.01	15.0	14.4	1.02	13.9	13.0	1.01
540	16.5	14.5	1.10	15.4	14.4	1.06	14.5	13.5	1.02
560	17.7	14.5	1.20	15.9	14.2	1.09	14.9	14.2	1.01
575	19.7	14.6	1.31	16.8	14.4	1.13	15.2	14.5	1.01

TABLE 4

Phase modulation fluorescence measurements on a  $2.5 \times 10^{-5}$  M ECMA-glycerol solution at 60 °C

$\lambda_{\text{obs}}$ (nm)	30 MHz			18 MHz		
	$\tau_{\phi}$	$\tau_m$	$m/\cos \phi$	$\tau_{\phi}$	$\tau_m$	$m/\cos \phi$
460	9.8	11.5	0.88	10.9	12.6	0.91
480	10.4	10.3	1.01	9.9	11.0	0.94
500	10.4	10.2	1.02	10.1	10.2	1.00
520	10.2	9.9	1.02	10.2	10.9	0.96
540	10.4	9.9	1.04	9.8	11.2	0.92
560	9.7	10.7	0.93	9.8	10.8	0.95
575	10.6	10.6	1.00	10.1	10.8	0.96

the mechanism of reorientation of solvent molecules around the F state was fast enough that fluorescence was emitted from the R state only, and thus the excited state process could not be detected in phase modulation measurements [3, 10].  $\tau_{\phi}$  and  $\tau_m$  were equal to  $10.5 \pm 0.6$  ns, and were now a measure of the lifetime of the excited state R.

### 3.2. Quantum yield measurements and determination of rate constants

The total fluorescence quantum yield  $Q_T$  was determined at 5, 21 and 60 °C (Table 5). By definition  $Q_T$  can be written (using the notation of Fig. 1) as

$$Q_T = \frac{k_{t,F}[F] + k_{t,R}[R]}{I_0} \quad (5)$$

where [F] and [R] represent the concentration of F states and R states respectively.

TABLE 5

Temperature variations in the fluorescence quantum yields, fluorescence lifetimes and deactivation rate constants of F and R states

$\theta$ (°C)	$Q_T$	$Q_F$	$\tau_F$ (ns)	$k_{f,F}$ ( $\times 10^7$ $s^{-1}$ )	$k_{FR}$ ( $\times 10^7$ $s^{-1}$ )	$k_{nr,F}$ ( $\times 10^7$ $s^{-1}$ )	$Q_R$	$\tau_R$ (ns)	$k_{f,R}$ ( $\times 10^7$ $s^{-1}$ )	$k_{nr,R}$ ( $\times 10^7$ $s^{-1}$ )
60	0.177	$\approx 0$					0.177	10.5	2.0	7.5
21	0.257	0.033	1.5	2.2	52.2	12.3	0.224	14.3	2.0	5.0
5	0.286	0.090	4.1	2.2	15.9	6.3	0.196	15.0	2.0	4.7

Under photostationary conditions we can write

$$\frac{d[F]}{dt} = 0 = I_0 - (k_{f,F} + k_{nr,F} + k_{FR})[F] \quad (6)$$

$$\frac{d[R]}{dt} = 0 = k_{FR}[F] - (k_{f,R} + k_{nr,R})[R] \quad (7)$$

Expressing eqn. (5) in terms of eqns. (6) and (7) it becomes

$$Q_T = k_{f,F}\tau_F + k_{FR}\tau_F k_{f,R}\tau_R \quad (8)$$

where

$$\tau_F = \frac{1}{k_{f,F} + k_{nr,F} + k_{FR}} \quad (9)$$

and

$$\tau_R = \frac{1}{k_{f,R} + k_{nr,R}} \quad (10)$$

The total fluorescence quantum yield can therefore be expressed as a sum of two terms. The first represents the F fluorescence quantum yield  $Q_F$ ; the second is not exactly the R fluorescence quantum yield since the R state is never directly photoexcited. Nevertheless we shall denote it  $Q_R$  since it is related to R emission. In addition, the product  $k_{FR}\tau_F$  represents the probability that an F state will be converted into an R state.

Phase modulation fluorescence data allow the resolution of the total fluorescence into its F and R components (Fig. 2) since  $\alpha$  was determined for various wavelengths. Consequently it is possible to calculate  $Q_F$  and  $Q_R$  at 21 °C and at 5 °C (Table 5). However, at 60 °C fluorescence was only emitted from the R state and eqn. (8) can then be written as

$$Q_T = Q_R = k_{FR}\tau_F k_{f,R}\tau_R$$

which implies that  $k_{f,F}$  is negligible relative to the product  $k_{FR}k_{f,R}\tau_R$ .

As  $\tau_F$  and  $Q_F$  were determined at 21 and 5 °C, the rate constant  $k_{f,F} = Q_F/\tau_F$  of fluorescence emission could be calculated at both temperatures. It

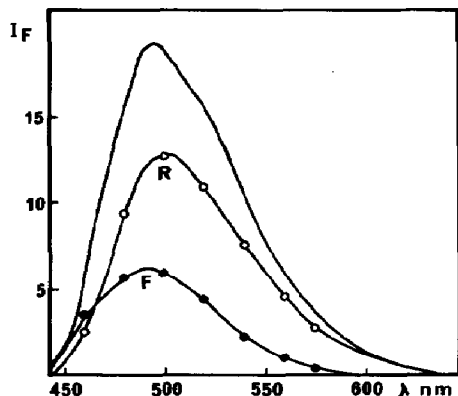


Fig. 2. Resolution of the total fluorescence of an ECMA-glycerol solution at 5 °C into its F and R components.

was equal to  $2.2 \times 10^7 \text{ s}^{-1}$  in both cases (Table 5). This rate constant changed with neither the temperature nor the viscosity of the medium and this completely agrees with the results of Birks [11]. Likewise, it seemed quite rational to us to assume that the rate constant  $k_{f,R}$  of the R state fluorescence emission did not significantly change with temperature [9, 12]. Nevertheless, the radiative rate constants do change with the refractive index  $n$  of the solvent used. In a water-ethanol solution ( $\theta = 21 \text{ }^\circ\text{C}$ )  $k_{f,R}$  has been determined as  $1.7 \times 10^7 \text{ s}^{-1}$  [9]. Consequently, its value in a glycerol solution ( $\theta = 21 \text{ }^\circ\text{C}$ ) can be calculated by using the equation

$$k_{f,R}(2) = \left(\frac{n_2}{n_1}\right)^2 k_{f,R}(1)$$

where 1 and 2 refer to the two different solvents [13]. We obtained  $k_{f,R} = 2 \times 10^7 \text{ s}^{-1}$ . The refractive index can be considered as constant for the glycerol solutions in the temperature range studied; consequently  $k_{f,R}$  was also constant. From eqn. (8) the solvent relaxation rate constant  $k_{FR}$  was calculated at 21 °C ( $k_{FR} = 52.2 \times 10^7 \text{ s}^{-1}$ ) and at 5 °C ( $k_{FR} = 15.9 \times 10^7 \text{ s}^{-1}$ );  $k_{nr,F}$  and  $k_{nr,R}$  were deduced from eqns. (9) and (10). All these results are listed in Table 5.

#### 4. Discussion

At 5 °C, the fluorescence emitted by an ECMA-glycerol solution arises from both F and R, F being the directly photoexcited state and R being the state that results from a reorientation of solvent molecules around the F state.

Let  $P$  be the deactivation probability of an F or R state, defined by the expression  $P = k\tau$  where  $k$  is the rate constant of the deactivation process concerned and  $\tau$  is the lifetime of the state considered.  $P_{FR}$ ,  $P_{nr,F}$ ,  $P_{f,F}$ ,  $P_{nr,R}$  and  $P_{f,R}$  can be calculated at 5 °C and at 21 °C (Table 6).

TABLE 6

Temperature variations in F and R state deactivation probabilities

$\theta$ (°C)	$P_{FR} = k_{FR}\tau_F$	$P_{f,F} = k_{f,F}\tau_F$	$P_{nr,F} = k_{nr,F}\tau_F$	$P_{f,R} = k_{f,R}\tau_R$	$P_{nr,R} = k_{nr,R}\tau_R$
60	0.84	$\approx 0$	0.16	0.21	0.79
21	0.78	0.03	0.18	0.29	0.71
5	0.65	0.09	0.26	0.30	0.70

Thus, of 100 F states directly photoexcited at 5 °C, 65 were converted into R states, 26 were deactivated by non-radiative transitions and 9 emitted fluorescence. Of the 65 R states produced, 19 emitted fluorescence and 46 were deactivated by non-radiative transitions.

At 21 °C, the process was qualitatively identical but quantitatively different. The temperature jump appeared in a large increase in  $k_{FR}$  and a relatively small increase in the radiationless rate constants  $k_{nr,F}$  and  $k_{nr,R}$ . Consequently, the lifetime of F decreased to a large extent while that of R decreased much less significantly. Therefore, of 100 F states directly photoexcited only 3 emitted fluorescence, 18 were deactivated by non-radiative transitions and 78 were converted into R states. Of the latter, 23 emitted fluorescence and 55 were deactivated by non-radiative transitions.

Raising the temperature to 60 °C results in an increase in  $k_{nr,F}$  and also in  $k_{FR}$ . Consequently, the lifetime of excited F was so short that the fluorescence probability  $P_{f,F}$  of the F state could be considered as negligible: the whole fluorescence emitted was ascribed to the R state. Using  $Q_R = 0.177$  and  $P_{f,R} = k_{f,R}\tau_R = 0.21$  it can be deduced from eqn. (8) that  $P_{FR} = 0.84$ . Consequently,  $P_{nr,R} = 0.79$  and  $P_{nr,F} = 0.16$  since  $P_{f,F}$  is assumed to be negligible. Thus, among 100 F states directly photoexcited at 60 °C, 84 were converted into R states of which 18 emitted fluorescence and 66 were deactivated by non-radiative transitions. The rate constant ratio  $k_{FR}/k_{nr,F}$  was calculated from the ratio of  $P_{FR}$  to  $P_{nr,F}$ . It was equal to 5.2 at 60 °C compared with 4.2 at 21 °C and 2.5 at 5 °C.

## Conclusions

In conclusion, an increase in temperature of an ECMA-glycerol solution induced an increase in the F and R radiationless transition rate constants as expected, and resulted in a more significant increase in the rate constant of the relaxation process. This strongly suggests that the rate determining step of this process is a solvent cage reorientation around photoexcited ECMA molecules.



## References

- 1 A. Marty and P. Viallet, *J. Photochem.*, 20 (1982) 213.
- 2 J. R. Lakowicz and H. Cherek, *Biochem. Biophys. Res. Commun.*, 99 (1981) 1173.
- 3 A. Marty, M. Bourdeaux, M. Dell'Amico and P. Viallet, *J. Photochem.*, 28 (1985) 71.
- 4 J. R. Lakowicz and A. Balter, *Biophys. Chem.*, 16 (1982) 99.
- 5 J. B. Birks, *J. Res. Natl. Bur. Stand., Sect. A*, 80 (1976) 389.
- 6 J. L. Cline Love, L. M. Upton and A. W. Ritter, *Anal. Chem.*, 50 (1978) 2059.
- 7 D. M. Jameson and G. Weber, *J. Phys. Chem.*, 85 (1981) 953.
- 8 R. D. Spencer and G. Weber, *J. Chem. Phys.*, 52 (1970) 1654.
- 9 A. Marty, Thesis "Doctorat d'Etat", Perpignan, France, 1984.
- 10 A. L. Campillo and T. M. Ordonez, *C. R. Acad. Sci. Paris*, 293 (1981) 271.
- 11 J. B. Birks, *Photophysics of Aromatic Molecules*, Wiley-Interscience, London, 1970, p. 312.
- 12 K. Kasama, K. Kikuchi, Y. Nishida and H. Kokubun, *J. Phys. Chem.*, 85 (1981) 4148.
- 13 R. A. Lampert, S. R. Meech, J. Metcalfe, D. Phillips and A. P. Schaap, *Chem. Phys. Lett.*, 94 (1983) 137.